

Review article of Phytochemical and Pharmacological activity of *Achyranthes aspera*, *Ficus glomerata* and *Leucas aspera* and Microbial activity test

Priyanka Biswas¹, Abhishek²

¹Department of Pharmacy,
Six Sigma Institute of
Technology and Science,
Jafarpur, Uttarakhand

²Assistant Professor,
Department of Pharmacy

Abstract:

The medicinal potential of medicinal plants is a well-known phenomenon from centuries. This review article is focus on the phytochemical, pharmacological, and microbial activity of three medicinal plants as *Achyranthes aspera* (Amaranthaceae), *Ficus glomerata* (Moraceae) and *Leucas aspera* (Lamiaceae). Its is rich source of bioactive compounds as alkaloids, flavonoids, tannins, saponins, phenolic compounds, and terpenoids. Its Various pharmacological activities have been reported as Hepatoprotective and antioxidant property DNA Damage protective activity immunomodulatory activity Antifungal activity Antimicrobial activity Antinociceptive activity antioxidant and cytotoxic activities. for the antimicrobial activity 80% methanol is use in which use, E.Coli, K.pneumoniae and S.typhi bacteria.

Keywords: Phytochemical, Pharmacological, *Achyranthes aspera*, *Ficus glomerata*, *Leucas aspera*, Microbial activity test

Corresponding Author: Priyanka Biswas[†], Department of Pharmacy , Six Sigma Institute of Technology and Science, Jafarpur, Uttarakhand

Copyright: © 2025 The Authors. Published by Vision Publisher. This is an open access article under the CC BY-NC-ND license (<https://creativecommons.org/licenses/by-nc-nd/4.0/>).

Introduction

Achyranthes aspera

The plants have rich source of phytochemicals in which have a wound healing and antioxidant property And antimicrobial properties It can help the reduce inflammation and infection of wound, cut, burn. *Achyranthes aspera* is commonly known as rough chaff tree in English. It is also known as Apamarga .this herb grow wild and Abundantly in India. India use the leave of this plant for healing wounds. *Achyranthes aspera* is reported various activity included Hepatoprotective(1), Cancer Chemo Preventive(2) ,Anti- inflammatory, anti- arthritic(3) ,thyroid stimulating, anti-

peroxidative(6), reproductive function(4), anti -bacterial(5), immunomodulatory(6), contraceptive (7),Saponian of this plants have phospholrylase activity on the heart root of these plants used for malarial fever hypertension asthma(8).

Phytochemical

Alkaloids, tannins, phenols, saponins, carbohydrates, steroids and glycosides are present in the achyranthes aspera(9).



Leave



Seed



Flower

Ficus glomerata

Ficus glomerata Roxb. syn. *F. racemosa* L. (Family: Moraceae) Gular in Hindi Cluster fig in English It is a medium-sized to large evergreen or occasionally deciduous tree and found throughout India and Southeast Asia. Its fruits are mixed with rice for making bread and used in several dishes. The bark, fruits and latex are used to treat anemia and gastrointestinal disorders like constipation, dysentery(10).

Phytochemical



Fruit



Leaf



Seed

The stem bark of *F. religiosa* have been reported to contain phytoconstituents of phenols, tannins, steroids, alkaloids and flavonoids β -sitosteryl-D-glucoside vitamin K n-octacosanol, methyl oleanolate, lanosterol, stigmasterol, lupen-3-one.(11) The major active constituent from the root bark *F. religiosa* was β -sitosteryl-D-glucoside, which exerts a peroral hypoglycemic action in fasting and alloxan-diabetic rabbits, and in pituitary-diabetic rats. The fruits contain 4.9% protein having the essential amino acids, isoleucine, and phenylalanine.(12) The seeds contain phytosterolin, β -sitosterol, and its glycoside, albuminoids, carbohydrate, fatty matter, coloring matter, caoutchoue 0.7–5.1%.(13) *F. religiosa* fruits contain flavonols namely kaempferol, quercetin, and myricetin (14) Leaves and fruits contain carbohydrate, protein, lipid, calcium, sodium, potassium, and phosphorus.(15) The aqueous extract of dried bark of *F. religiosa* has been reported to contain phytosterols, flavonoids, tannins, furanocoumarin derivativesnamely bergapten and begaptol(16)

Pharmacological property

Hepatoprotective and antioxidant property-The methanol extract of the bark when given orally along with CCl₄ at the doses of 250 and 500 mg/kg body weight showed a significant reversal of these biochemical changes towards the normal when compared to CCl₄-treated control rats in serum, liver and kidney(17)

DNA Damage protective activity

Green fruit of *Ficus glomerata* is show protective oxidative DNA Damage(18)

Immunomodulatory activity

Bark and Fruit of *Ficus glomerata* is show phagocytic effect on human neutrophils (19)

Leucas aspera

Leucas aspera (Lamiaceae) is a small herbaceous, erect plant with free blooming nature, which grows as a weed on waste lands and roadsides all over India. It is pungently aromatic and commonly used as antipyretic herb in south India. In traditional medicine it is indicated for cold, cough, painful swellings and chronic skin eruptions. The plant was previously evaluated pharmacologically for its anti-inflammatory, analgesic and protective effects against cobra venom poisoning (20)

Phytochemical

L. aspera revealed presence of triterpenoids in entire plant(21). Whole plant is reported to contain oleanolic acid, ursolic acid and 3-sitosterol(22)Aerial parts are reported to contain nicotine(23), sterols(24),two new alkaloids (compound A m.p. 61-2°, α -sitosterol and β -sitosterol) (m.p. 183-4°), reducing sugars (galactose), glucoside (230-1°)(25),diterpenes (leucasperones A and B, leucasperols A and B, isopimarane glycosides (leucasperosides A, B and C), together with other compounds like asperphenamate, maslinic acid, (-)-isololiolide, linifolioside(26),] nectandrin B, meso-dihydroguaiaretic acid, macelignan, acacetin, apigenin 7-O-[6'-O-(p-coumaroyl)-3-D-glucoside], chrysoeriol, apigenin, erythro-2-(4-allyl-2,6-dimethoxyphenoxy)-1-(4-hydroxy-3-methoxyphenyl)propan-1-ol, myristargenol B, and machilin C, (-)-chicanine, (7R,8R)- and (7S,8S)-licarin A(27), Among the 25 compounds identified from the leaf volatiles, u-farnesene (26.4%), x-thujene (12.6%) and menthol (11.3%) were the major constituents. The flower is reported to contain 10 compounds; among them amyl propionate (15.2%) and isoamyl propionate (14.4%) were dominant(28),Seed is reported to contain palmitic acid (6.25%), stearic acid (2.84%), oleic acid (42.07%), linoleic acid (48.11%), and linolenic acid (0.65%). The unsaponifiable fraction contained 3-sitosterol and ceryl alcohol(29),Shoot contained novel phenolic compounds (4-(24-hydroxy-1-oxo-5-n-propyltetracosanyl)-phenol)(30),aliphatic ketols (28-hydroxypentatriacontan-7-one, 7-hydroxydotriacontan-2-one)(31),long-chain compounds (1-hydroxytetracontan-4-one, 32-methyltetracontan-8-ol)(32), nonatriacontane,5-acetoxytetracontane, β -sitosterol(33) and dotriacontanol, Leucolactone (I), isolated from the root of *L. aspera* have been characterized as 3,3,16c-dihydroxyoleanan-28-1,3-olide(34).



Seed



Plant

Pharmacological activity

Antifungal activity

In an in vitro study of chloroform and ether extracts that revealed antifungal activity against *Trichophyton* and *Microsporum gypseum* L. *aspera*, minimum inhibitory concentration was observed to be at 5mg/mL(35).

Activity of *Leucas aspera* flowers on antimicrobial

The methanolic extract of *L. aspera* flowers, its fractions and the expressed flower juice all showed good antibacterial activities for methanolic extracts and methanolic fractions which were found to have maximum activities for the alkaloidal residues(36).

Anti-nociceptive, antioxidant and cytotoxic activities

The ethanolic extract of *L. aspera* root caused a significant inhibition in acetic acid-induced writhing in mice at doses of 250 and 500 mg/kg. The extract showed significant free radical scavenging activity with an IC₅₀ of 8 µg/ml. The extract was showed significant lethality to brine shrimp(37).

Antimicrobial activity Test

MATERIALS AND METHODS

Collection of medicinal Plants

Three medicinal plants were selected and their parts(only full grown and matured parts) during months of april and may.

Preparation of alcohol extract

the collected medicinal plant were cleaned and dried under shade. The dry plant material was converted into fine powder. 500 mg of dry powder was extract with 80% alcohol at 60-80o C by using soxhlet apparatus. the extraction was continued for 24 hours. the alcoholic extract was then filtered and kept in oven at 50oC for 24 hours to evaporate the alcohol from it. A dark brown residue was obtained. the solid fractions was redissolve in dimethyl formamide(DMF) and their microbial efficiency were noted. DMF was a inert organic solvent.

Selection of microorganism

Escherichia coli bacteria, *Klebsiella pneumoniae* bacteria and *Salmonella Typhi* were use for the antimicrobial activity. *E.Coli*, *K.pneumoniae* and *S.typhi* were used in the antimicrobial screening for 24 hours cultures .

Microbial test

5% w/v test solution of each extract was prepared by dissolving 250mg of each extract separately in 5ml of sterile dimethyl formamide (DMF). Nutrient agar medium was prepared and sterilized by an autoclave. In an aseptic room, they poured unto sterile petridishes to a uniform depth of 4mm and then allowed to solidify at room temperature. After solidification, the test organism were inoculated with the help of a sterile swab soaked in a bacterial culture of suspension. Thus provides the uniform surface growth of bacterium and used for antibacterial sensitivity studies. Then the sterile filter paper discs (6mm) containing sample (100ul) were immersed in plant extracts and was placed over the solidified agar in such a way that there is no overlapping of zone of inhibition (38). Plates were kept at room temperature for half an hour for the diffusion of the sample into the agar media. The organisms inoculated petridishes were incubated at 37oC for 48 hours. After the incubation period is over, the zone of inhibition produced by the sample with different organisms in different plates were measured and recorded immediately by using a zone reader (39)

Results and Discussion:

the extracts of *A. aspera*, *F. glomerata*, *L. aspera* is conducted to analyze the antimicrobial activity of the *E.coli*, *K. pneumoniae* and *S.typhi* bacteria. zone of inhibition of *A.asper* show 7mm, 9mm, and 8mm, *F.glomerata* is show 7mm, 15mm and 8mm by zone of inhibition, *L.asper* is show 15mm, 14mm and 11mm by zone of inhibition. All zone of inhibition is measured and compared with the standard values. There is no zone of inhibition in the control (DMF)

against *E.coli*, *K. Pneumoniae*, and *S. typhi*. Depending upon the values of the measured diameter of complete inhibition of the circle containing the disc, in millimeter, the antibacterial activity can be categorized into the following types: >12mm zone of inhibition-highly sensitive, 9-12 mm zone inhibition-moderately sensitive, 6-9 mm zone of inhibition-less sensitive and <6 mm zone of inhibition-resistant (15). Thus the present study, on record are sensitivity more than 12mm zone of inhibition shown by t-extract of *F.glomerta* against *K.Pneumoniae* (15mm), *L.aspera* against *E.coli* (15mm) and *K. Pneumoniae* (14mm). Antimicrobial activity are expressed through the extracts as if possibly due to certain antimicrobial substances contents present within.

Conclusion

Achyranthes aspera, *Ficus glomerata*, and *Leucas aspera* are very rich in phytochemical diversity and pharmacological potency. The bioactive compounds in these plants have a variety of therapeutic properties such as anti-inflammatory, analgesic, antioxidant, anticancer, and antimicrobial activities. The microbial activity tests results show that these plants have strong antimicrobial activity and can be used in the development of drugs to overcome the drug-resistant pathogens. Despite the positive findings, more research work, especially clinical trials, would be necessary to fully exploit these plants for modern medicine. With their rich chemical composition and multifaceted activities, the plants may become a treasure in the development of new therapeutic agent.

Reference

1. AR Bafna, and SH Mishra. (2004). Effect of methanol extract of *Achyranthes aspera* Linn. on rifampicin-induced hepatotoxicity in rats. *Ars Pharmaceutica* 45:343–351.
2. A Chakraborty, A Brantner, T Mukainaka, Y Nobukuni, M Kuchide, T Konoshima, H Tokuda, and H Nishino. (2002). Cancer chemopreventive activity of *Achyranthes aspera* leaves on Epstein-Barr virus activation and two-stage mouse skin carcinogenesis. *Cancer Lett* 177:1–5.
3. AB Gokhale, AS Damre, KR Kulkarni, and MN Saraf. (2002), Preliminary evaluation of anti-inflammatory and anti-arthritic activity of *S. lappa*, *Achyranthes speciosa* and *Achyranthes aspera*. *Phytomedicine* 9:433–437.
4. Tahiliani P, Kar A. elevates the *Achyranthes aspera* for thyroid hormone levels and decreases hepatic lipid peroxidation in male rats. *J Ethnopharmacol.* 2000 Aug;71(3):527-32.
5. K Sandhyakumary, RG Bobby, and M Indira. (2002), Impact of feeding ethanol extracts of *Achyranthes aspera* Linn. on reproductive functions in male rats. *Indian J Exp Biol* 40:1037–1039
6. DK Verma, SK Singh, and V Tripathi. (1997). A rare antibacterial activity of *Achyranthes aspera* Linn. *Indian Drugs* 34:32–35..
7. G Rao. (2002). Immunomodulatory activity of *Achyranthes aspera* on the elicitation of antigen specific murine antibody response. *Int J Pharmacog* 40:175–178.
8. V Wadhwa, MM Singh, DN Gupta, C Singh, and VP Kamboj. (1986), Contraceptive and hormonal properties of *Achyranthes aspera* in rats and hamsters. *Planta Med* 3:231–233.
9. Das AK, Bigoniya P, Verma NK, Rana AC. Gastroprotective effect of *Achyranthes aspera* Linn. leaf on rats. *Asian Pac J Trop Med*. 2012 Mar.
10. Verma AR, Vijayakumar M, Rao CV, Mathela CS. In vitro and in vivo antioxidant properties and DNA damage protective activity of green fruit of *Ficus glomerata*, *Food Chem Toxicol*, 2010 Feb.
11. Chandrasekar SB, Bhanumathy M, Pawar AT, Somasundaram T. Phytopharmacology of *Ficus religiosa*. *Pharmacogn Rev.* 2010 Jul;4(8):195-9. doi: 10.4103/0973-7847.70918. PMID: 22228961; PMCID: PMC3249921.
12. Sheetal A, Bagul MS, Prabha M, Rajani M. Evaluation of free radicals scavenging activity of an Ayurvedic formulation, panchvankala. *Indian J Pharm Sci.* 2008.

13. Oliver bever B. Oral hypoglycaemic plants in West Africa. J Ethnopharmacol, 1977.
14. Khare CP. Encyclopedia of Indian medicinal plants. Berlin Heidelberg, New York, Springer-Verlag,2004.
15. Bushra S, Farooq A. Flavonols (kaempeferol, quercetin, myricetin) contents of selected fruits, vegetables and medicinal plants, Food Chem. 2008.
16. Ruby J, Nathan PT, Balasingh J, Kunz TH. Chemical composition of fruits and leaves eaten by short- nosed fruit bat, *Cynopterus sphinx*. J Chem Ecol. 2000.
17. Channabasavaraj KP, Badami S, Bhojraj S. Hepatoprotective and antioxidant activity of methanol extract of *Ficus glomerata*. J Nat Med. 2008 Jul.
18. Verma AR, Vijayakumar M, Rao CV, Mathela CS. In vitro and in vivo antioxidant properties and DNA damage protective activity of green fruit of *Ficus glomerata*. Food Chem Toxicol. 2010 Feb.
19. Heroor S, Beknal AK, Mahurkar N. Immunomodulatory activity of methanolic extracts of fruits and bark of *Ficus glomerata* Roxb. in mice and on human neutrophils. Indian J Pharmacol. 2013 Mar-Apr;45(2):130-5. doi: 10.4103/0253-7613.108287. PMID: 23716887; PMCID: PMC3660923.
20. Ravindran R, Juliet S, Sunil AR, Kumar KG, Nair SN, Amithamol KK, Shynu M, Rawat AK, Ghosh S. Eclosion blocking effect of ethanolic extract of *Leucas aspera* (Lamiaceae) on *Rhipicephalus* (*Boophilus*) *annulatus*. Vet Parasitol. 2011 Jun 30;179(1-3):287-90. doi: 10.1016/j.vetpar.2011.02.021. Epub 2011 Mar 4. PMID: 21440993.
21. Kamat M, Singh TP. Preliminary chemical examination of some compounds in the different parts of the genus *Leucas*. Geobios. 1994.
22. Chaudhury NA, Ghosh D. Insecticidal plants: Chemical examination of *Leucas aspera*. J Indian Chem Soc. 1969.
23. Mangathayaru K, Thirumurugan D, Patel PS, Pratap DV, David DJ, Karthikeyan J. Isolation and identification of nicotine from *leucas aspera* (willd) Indian J Pharm Sci. 2000.
24. Khaleque A, Huq ME, Huq MS, Mansoor MH. Chemical investigations on *Leucas aspera*. I. Isolation of compound-A, 3-sitosterol and et-sitosterol from the aerial parts. Scientifi c Res. 1970.
25. Chatterjee SK, Majumdar DN. Chemical investigation of *Leucas aspera*. J Inst Chem. 1969.
26. Sadhu SK, Okuyama E, Fujimoto H, Ishibashi M. Diterpenes from *Leucas aspera* inhibiting prostaglandin-induced contractions. J Nat Prod. 2006.
27. Sadhu SK, Okuyama E, Fujimoto H, Ishibashi M. Separation of *Leucas aspera*, a medicinal plant of Bangladesh, guided by prostaglandin inhibitory and antioxidant activities. Chem Pharm Bull (Tokyo) 2003.
28. Kalachaveedu M, Ghosh A, Ranjan R, VedamVenkat K. Volatile constituents of *Leucas aspera* (Willd.) J Essent Oil Res. 2006.
29. Jam MP, Nath HB. Examination of the component fatty acids of the oil from the seeds of *Leucas aspera*. Lab Dev. 1968.
30. Badami RC, Patil KB. Minor seed oils.X: Physico-chemical characteristics and fatty acid composition of seven minor oils. J Oil Technol Assoc India. 1975.
31. Misra TN, Singh RS, Pandey HS, Singh S. A novel phenolic compound from *Leucas aspera* Spreng. Indian J Chem Br. 1995.
32. Misra TN, Singh RS, Prasad C, Singh S. Two aliphatic ketols from *Leucas aspera*. Phytochemistry. 1992.
33. Misra TN, Singh RS, Pandey HS, Singh S. Long-chain compounds from *Leucas aspera*. Phytochemistry. 1992.
34. Pradhan B, Chakraborty D, Subba G. A triterpenoid lactone from *Leucas aspera*. Phytochemistry. 1990.

35. Thakur DK, Misra SK, Choudhuri PC. In vitro trials of plant extracts and chemicals for their antifungal activity. *Indian J Animal Health*. 1987;26:31–5.
36. Mangathayaru K, Lakshmikanth J, Shyam Sundar N, Swapna R, Grace XF, Vasantha J. Antimicrobial activity of *Leucas aspera* flowers. *Fitoterapia*. 2005;76:752–4. doi: 10.1016/j.fitote.2005.08.009.
37. Rahman MS, Sadhu SK, Hasan CM. Preliminary antinociceptive, antioxidant and cytotoxic activities of *Leucas aspera* root. *Fitoterapia*. 2007;78:552–5. doi: 10.1016/j.fitote.2006.06.018.
38. Maryzzella, J.C. and practical , A.H., *J.Am.Pharm. Assoc.*, 47,471 (1958).
39. Marian, T.G., Murthy, P.N. Ranganatham, P.Hymete, A. and Daka, K. *The Eastern Pharmacist*, Vol 36, No.33,131 (1993)