

## Antimicrobial Properties of Ethanolic and Other Components Chromatographic Analysis of *Artemisia annua* Leaf Extract and Its Functional Groups

Rabab J.H. Al Hasseny<sup>1</sup>, Abbas K. Al-Mansoori<sup>2</sup>, Toqa M.A. Mhayyal<sup>3</sup>, Fatima ABD-AL-RADHIA<sup>4</sup>

<sup>1</sup>Medical Microbiology,  
Department of Health  
Food and Nutrition,  
College of Food Science,  
Al-Qasim Green  
University, Iraq

<sup>2</sup>Department of Genetic  
Engineering, College of  
Biotechnology, Al-Qasim  
Green University, Iraq

<sup>3,4</sup>AL-Qasim Green  
University, Faculty of  
Biotechnology,  
Department of Medical  
Biotechnology, Iraq

### Abstract:

Numerous studies have demonstrated that the essential oil and extracts of *Artemisia annua* has antibacterial characteristics, rendering them efficient against many bacteria and fungi. Some researchers think that the plant's ability to kill both Gram-positive and Gram-negative bacteria is due to the bioactive chemicals it contains, including as artemisinin and terpenes. Using chromatographic analysis of functional groups and ethanolic *Artemisia annua* leaf extract, this study intends to assess the antibacterial capabilities of the plant. The results of the GC-MS analysis proved the presence of the compounds, which are 9-Octadecenoic acid, (-)-delta-cadinene, Tetraeneurin E, 1-tert-Butyl-4,4-diphenylpiperidine, Lucenin-2, Germacrene D-4-ol, 4-(2,3-Diphenylcyclopropyl)phenol, Farnesene, Tetracosahexaenoic acid, beta-Caryophyllene oxide, and Isochinol. According to (methanol, Ethyl acetate, and Ethanol fraction), Ampicillin (AMP) and Cefotaxime (CTX). Recorded (28.00±0.45, 18.00±0.28, 23.00± 0.40, 33.01±0.51 and 30.00±0.49) respectively in *Staphylococcus aureus*. While recorded (22.91±0.81, 14.09±0.23, 24.00±0.41, 29.00±0.48 and 26.98±0.45) for *Escherichia coli*. Record (24.19±0.38, 18.09±0.27, 21.26±0.29, 31.08±0.50 and 28.16±0.49) *Streptococcus pyogenes*. While (21.95±0.38, 25.96±0.42, 27.00±0.44, 27.95±0.48 and 23.00±0.45) for *Staphylococcus epidermidis*. The metabolites of *Artemisia annua* exhibited significant activity against *Staphylococcus aureus* (28.00±0.45).

**Keywords:** Chromatographic Analysis, Antimicrobial Properties, *Artemisia annua*, Functional Groups



**Corresponding Author:** Rabab J.H. Al Hasseny<sup>†</sup>, Medical Microbiology, Department of Health Food and Nutrition, College of Food Science, Al-Qasim Green University, Iraq

**Copyright:** © 2025 The Authors. Published by Vision Publisher. This is an open access article under the CC BY-NC-ND license (<https://creativecommons.org/licenses/by-nc-nd/4.0/>).

# INTRODUCTION

Herbs and shrubs of the genus *Artemisia* are mostly found in semiarid (steppe climate) areas. According to what is known from published accounts, this genus contains about 500 species. As a result of the specimens' many geographic origins, it exhibits a high degree of morphological and phytochemical heterogeneity. Furthermore, the taxonomic and evolutionary relationships of the genus *Artemisia* are the subject of numerous investigations. Sesquiterpenes, terpenoids, and monoterpenes in essential oils are among the biologically active components of the *Artemisia* plant that give it scientific significance [1, 2]. A broad range of intriguing biological actions are associated with these types of phytochemicals. Emergence and spread of fungal and bacterial infectious illnesses have become a major cause of death for all forms of life on Earth. The primary weapon in the battle against infectious diseases is the antibiotic. It is becoming increasingly difficult to treat these infections due to the rise of multidrug-resistant microbes, which is having a major impact on healthcare costs. Oxidative stress, which occurs when the production of ROS outpaces their elimination, is another major threat to human health. Cell membrane oxidative damage, destruction of various biological substances, and numerous serious diseases are all results of this occurrence [3]. As a result, developing novel, potent compounds to combat free radicals and drug-resistant bacteria is of the utmost importance. To get around these issues, researchers have been focusing more on using natural components to create novel medications that are both effective and safe. A gift from Mother Nature, medicinal and aromatic plants (MAP) enable humans to live long, healthy lives free from illness. Natural medicine has a long and storied history of use in human and veterinary medicine, dating back to the dawn of recorded history. An increasing number of people are considering using natural medicines instead of synthetic ones because of the potential side effects and high cost of the former. Natural plants belonging to the Asteraceae family have a long history of widespread use in popular cuisine around the world due to their high nutritional content [4-7]. Moroccan tea is a popular herbal tea among diabetics. It is often used as an appetizer, condiment, spice, or in salads. They have chemical components that can be used to create medications that can compete with synthetic pharmaceuticals; in fact, some of these compounds have already been given the green light for clinical use [8-10]. The objective of this research is to analyze the antimicrobial characteristics of *Artemisia annua* leaf extract, its functional groups, and various ethanolic fractions.

## MATERIALS AND METHODS

**Methods for Extracting Ethanol from Herbs and Other Plant Materials** The plant parts used in this study were the leaves and aerial portions of *Artemisia annua* L. plants, which are native to Hillah City in Iraq. The plant's aerial parts were cut into medium-sized pieces and then dried in a chamber at room temperature for two weeks at the University of Babylon. The final product was dried using anhydrous sodium sulfate ( $\text{Na}_2\text{SO}_4$ ), then put in amber worm bottles and kept at 4 °C until it was time for testing. After drying the extract, it was preserved at a temperature of -18 degrees Celsius until needed.

### Chemical Composition Analysis with GC-MS

*Artemisia annua*'s chemical composition was determined using GC-MS analysis. In order to identify and quantify the components, 1  $\mu\text{L}$  was passed through a gas chromatography–mass spectrometer (GC-MS–TQ8040 NX, brand Shimadzu, Tokyo, Japan) fitted with an apolar capillary column (RTxi-5 Sil MS-30m  $\times$  0.25 mm ID  $\times$  0.25  $\mu\text{m}$ ). Starting at 50 °C for two minutes, the oven temperature was gradually increased at a rate of 5 °C per minute until it reached 160 °C. After that, it was ramped up to 280 °C for two minutes. It is worth mentioning that the analysis was conducted for 50 minutes using 1 mL/min of high-purity helium as the carrier gas, with a split ratio of 1:20. A temperature of 280 °C was maintained for the detector, while an injection temperature of 250 °C was maintained. This experiment used a spectral range of  $m/z$  40–650, an electron ionization energy of 70 eV, and an ion source temperature of 200 °C. The volume of the sample that was injected was 1  $\mu\text{L}$ . We used the NIST-MS Search Version 2.0 application to compare the mass spectra of the chemical compounds in the sample to those in the MS library. This

allowed us to identify the chemicals in the sample. In addition, the Adams database served as a reference for comparing their Kovats index..

### Antimicrobial Activity Determining the Minimum Inhibitory Concentration (MIC)

Traditional harmful bacterial strains like *S. aureus*, *E. coli*, *Streptococcus pyogenes*, and *S. epidermidis* have already undergone experimental screening. For twenty-four hours at 37 degrees Celsius, they were actually cultured in a petri dish with 10 milliliters of sterile nutritional broth. Only one day was all the incubation time for the plates at 37°C. In the experiment, an optical microscope was utilized to confirm the form of the bacterium. Meanwhile, sterile saline was used to disperse the number of colonies actually belonging to each strain. This was then adjusted in the lab so that it matched the turbidity of the 0.5 McFarland standard, which was made on Mueller-Hinton agar and contained 1.5 x 10<sup>8</sup> colony-forming units per milliliter.

### Data Analysis by Statistic

The data is shown as the mean plus or minus the standard deviation. A program called GraphPad Prism 9 was used to conduct the analysis. We employed analysis of variance (ANOVA) and Tukey's extremely significant difference test to compare the results. When the p-value was lower than 0.05, it was assumed that there was a statistically significant difference.

## RESULTS AND DISCUSSION

According to the World Health Organization, traditional medicine is currently practiced by 80% of the population in developed countries. An important genus of herbs, *Artemisia* is officially recognized as *Asteraceae annua* L. Species identified in this group are mostly associated with northern temperate climates, where the average yearly rainfall is between zero and fifty centimeters [11, 12]. Sweet wormwood, or *Artemisia annua* L., is a member of the *Asteraceae* family of plants that grows wild mostly in Asia, more specifically in Korea, Japan, and China. It has also spread to other nations, where it is now grown. The results of the GC-MS analysis proved the presence of the compounds, which are  $\nu$ 9-Octadecenoic acid, (-)-delta-cadinene, Tetraneurin E, 1-tert-Butyl-4,4-diphenylpiperidine, Lucenin-2, Germacrene D-4-ol, 4-(2,3-Diphenylcyclopropyl)phenol, Farnesene, Tetracosahexaenoic acid, beta-Caryophyllene oxide, and Isochinol.

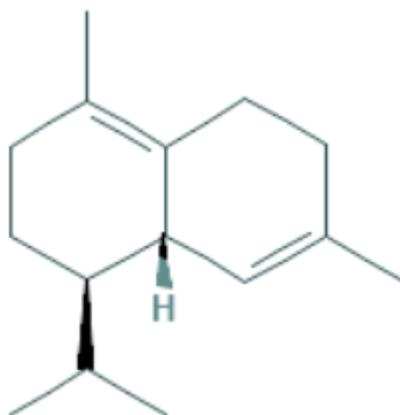
### 9-Octadecenoic acid



MF: C<sub>18</sub>H<sub>34</sub>O<sub>2</sub>

MW: 282.5 g/mol

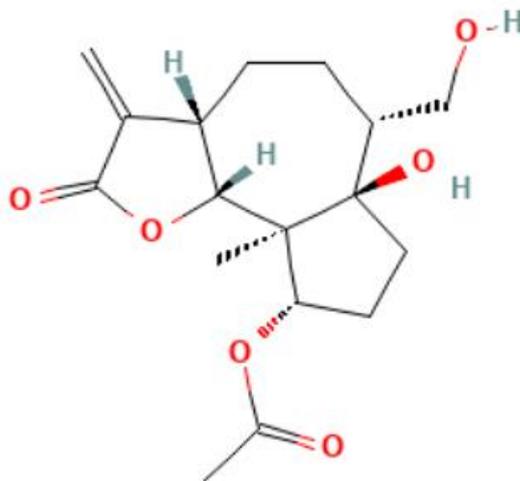
### (-)-delta-cadinene



MF: C<sub>15</sub>H<sub>24</sub>

MW: 204.35 g/mol

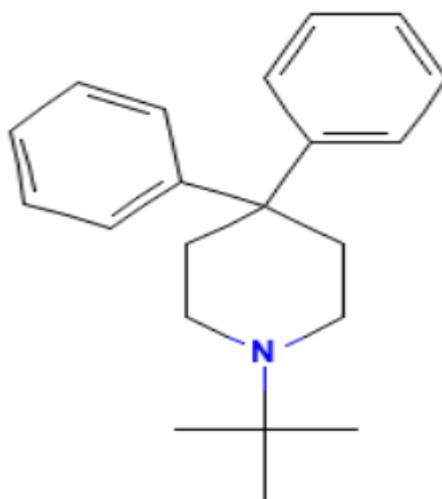
**Tetraneurin E**



**MF: C<sub>17</sub>H<sub>24</sub>O<sub>6</sub>**

**MW: 324.4 g/mol**

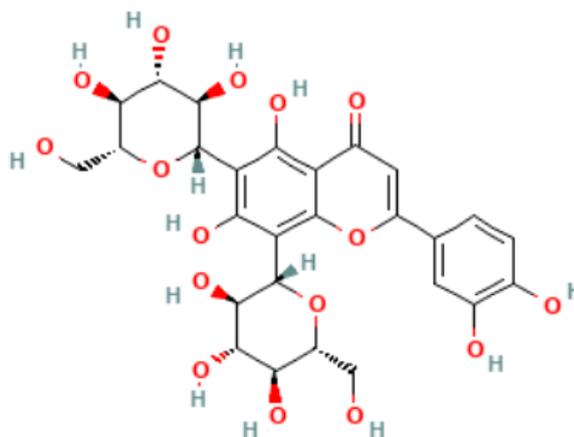
**1-tert-Butyl-4,4-diphenylpiperidine**



**MF: C<sub>21</sub>H<sub>27</sub>N**

**MW: 293.4 g/mol**

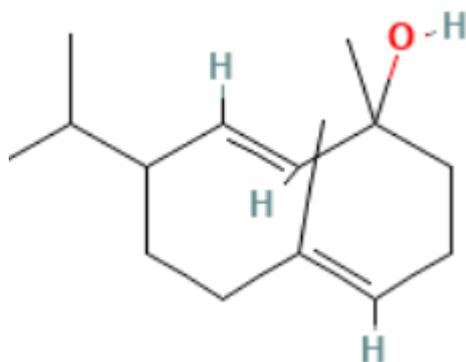
**Lucenin-2**



**MF: C<sub>27</sub>H<sub>30</sub>O<sub>16</sub>**

**MW: 610.5 g/mol**

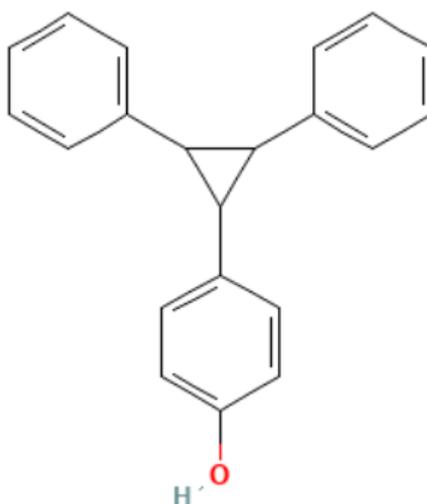
**Germacrene D-4-ol**



**MF: C<sub>15</sub>H<sub>26</sub>O**

**MW: 222.37 g/mol**

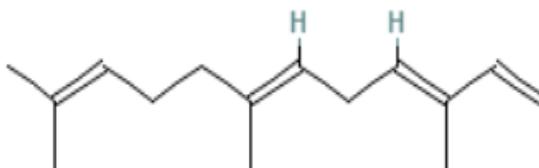
**4-(2,3-Diphenylcyclopropyl)phenol**



**MF: C<sub>21</sub>H<sub>18</sub>O**

**MW: 286.4 g/mol**

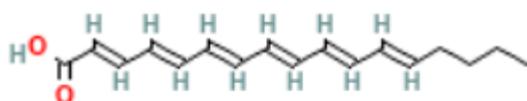
**Farnesene**



**MF: C<sub>15</sub>H<sub>24</sub>**

**MW: 204.35 g/mol**

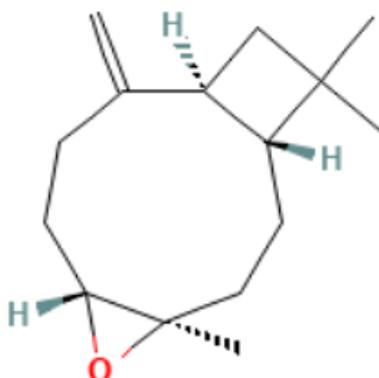
**Tetracosahexaenoic acid**



**MF: C<sub>24</sub>H<sub>36</sub>O<sub>2</sub>**

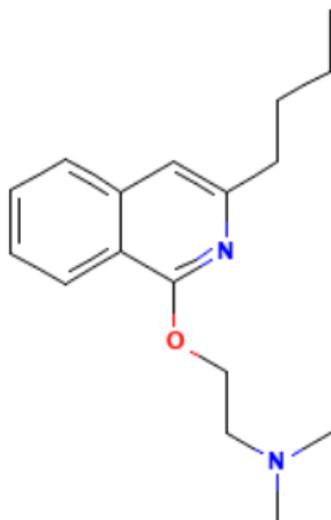
**MW: 356.5 g/mol**

**beta-Caryophyllene oxide**

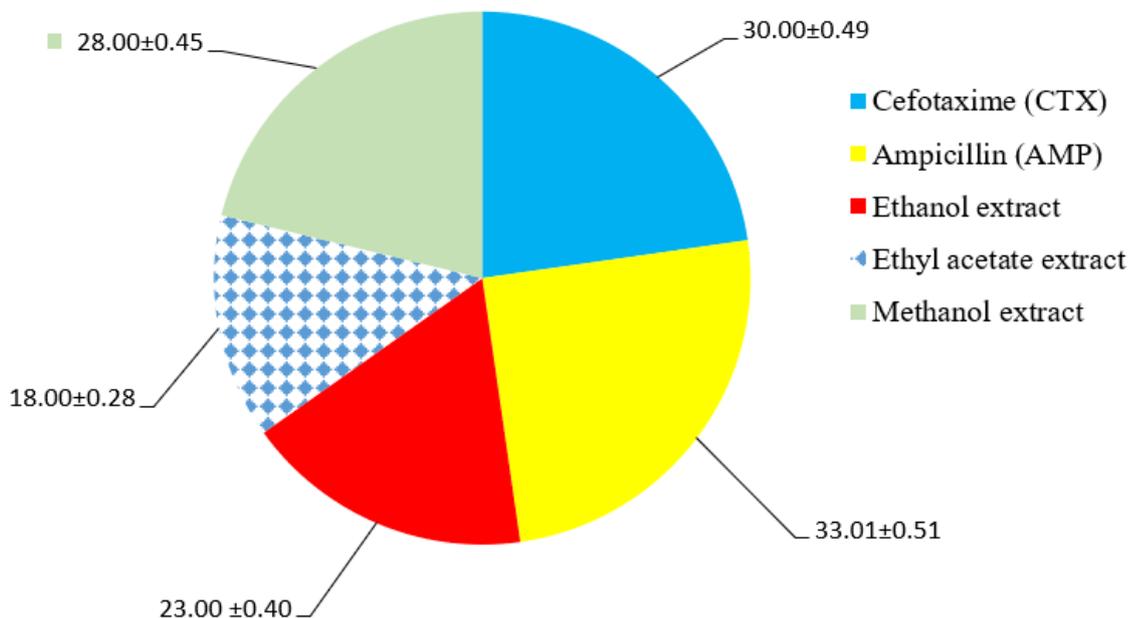


**MF: C<sub>15</sub>H<sub>24</sub>O**

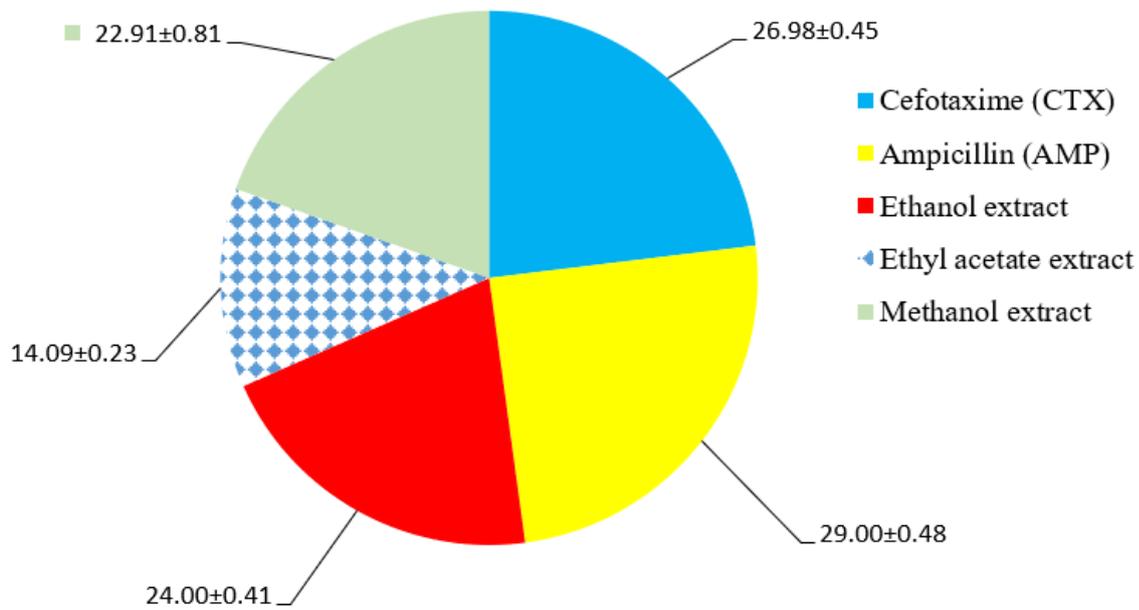
**MW: 220.35 g/mol**



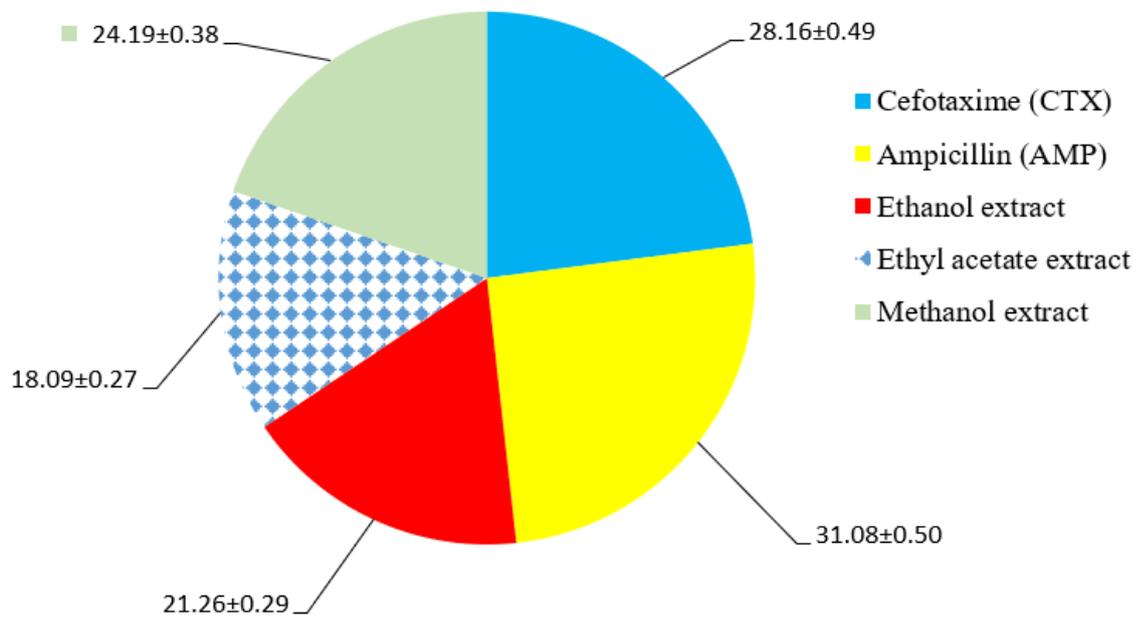
In vitro tests were conducted on four distinct microbial species to determine the antibacterial activity of leaf extracts of *Artemisia annua*. Cefotaxime (CTX) and Ampicillin (AMP) are based on the (methanol, ethyl acetate, and ethanol portion). *Staphylococcus aureus* tested positive for (28.00±0.45, 18.00±0.28, 23.00±0.40, 33.01±0.51 and 30.00±0.49) respectively. Although the results for *Escherichia coli* were recorded as (22.91±0.81, 14.09±0.23, 24.00±0.41, 29.00±0.48, and 26.98±0.45). The *Streptococcus pyogenes* record includes values of (24.19±0.38, 18.09±0.27, 21.26±0.29, 31.08±0.50, and 28.16±0.49). For *Staphylococcus epidermidis*, the values are (21.95±0.38, 25.96±0.42, 27.00±0.44, 27.95±0.48, and 23.00±0.45). *Staphylococcus aureus* was significantly inhibited by the metabolites of *Artemisia annua* (28.00±0.45).



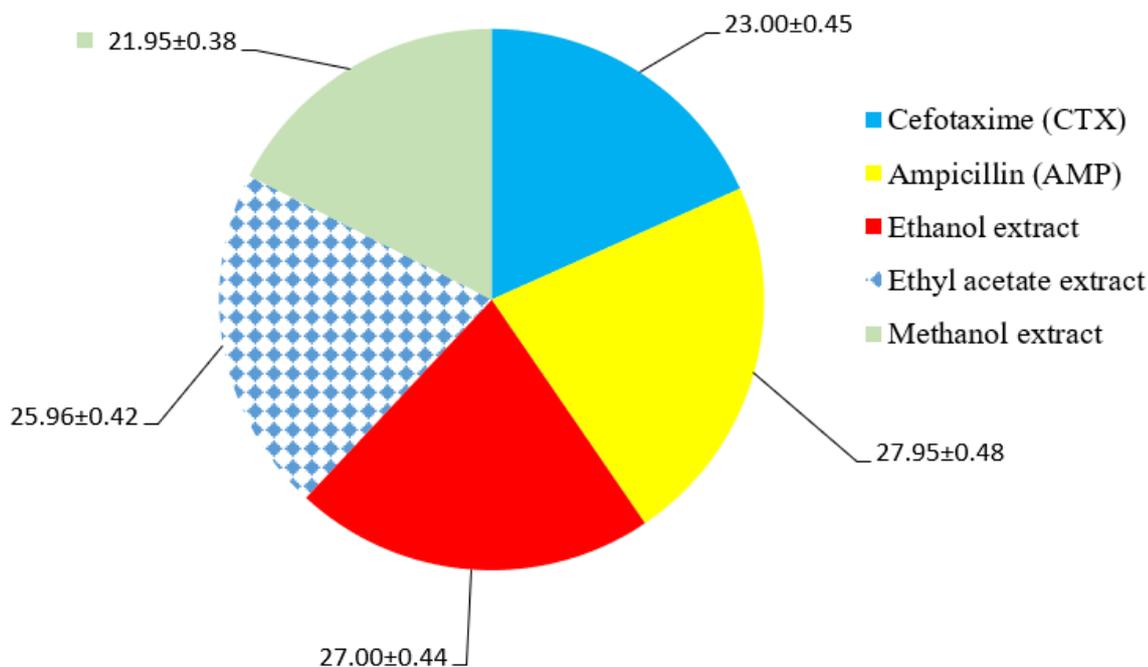
**Figure 1. Antibacterial activity of bioactive compounds of *Artemisia annua* and standard antibiotics against *Staphylococcus aureus***



**Figure 2. Antibacterial activity of bioactive compounds of *Artemisia annua* and standard antibiotics against *Escherichia coli***



**Figure 3. Antibacterial activity of bioactive compounds of *Artemisia annua* and standard antibiotics against *Streptococcus pyogenes***



**Figure 4. Antibacterial activity of bioactive compounds of *Artemisia annua* and standard antibiotics against *Staphylococcus epidermidis***

Metabolites with antibacterial characteristics are produced by multiple *Artemisia* species. The ethanolic extract also had a high concentration of chlorogenic acid, which was discovered in a tall variety of Asteraceae. Recent studies have shown that chlorogenic acid can induce cell death by binding to the outer membrane [13-15], disrupting it, decreasing the intracellular potential, and releasing macromolecules from the cytoplasm. Disappointing patient outcomes and increased healthcare costs are the results of antibiotic resistance, a worldwide problem that is only getting worse. The increasing resistance to commonly prescribed antibiotics like tetracycline and sulfamethoxazole has made it increasingly challenging to combat severe bacterial infections caused by Enterobacteriaceae, such as *Escherichia coli*, and other Gram-negative bacteria. This is especially true given the limited therapeutic options available for multidrug-resistant strains and the lack of new drug development. Additionally, resistance to ampicillin, chloramphenicol, gentamicin, ciprofloxacin, trimethoprim/sulfamethoxazole, and tobramycin has significantly increased during the past few years [16-19]. The use of traditional medicinal plants to treat a wide range of illnesses is becoming increasingly popular around the world. It is worth mentioning that *Artemisia* is one of the biggest genera in the Asteraceae family and is found all over the northern hemisphere. To enhance treatment procedures and perhaps supplant conventional medicines, medicinal plants can be combined with active pharmacological and nutritional components [20, 21].

Essential oils extracted from plants have grown in popularity as a result of their low toxicity and widespread use as treatments for serious illnesses. To that end, we tested the antioxidant and antibacterial capabilities of essential oils extracted from the Mediterranean-dwelling *Artemisia annua* L. Thanks to our EO extraction process, we were able to achieve a yield of about 0.51%. We found that the yield varies depending on when it is harvested, where the seeds are grown, how the leaves or flowers are used, and the location of the culture. Importantly, chemotype, plant part, genotype, drying circumstances, harvest season, geographic location, fertilizer, soil pH, and extraction process are some of the factors that can affect chemical compound variance [22-25]. There were a lot of oxygenated monoterpenes (55.35 percent) and hydrocarbon sesquiterpenes (13.43%) in the EOAA analysis. While noting the lack of oxygenated monoterpenes, a smaller quantity of oxygenated monoterpenes (1.62%) was found. Essential oils (EOs) derived from the *Artemisia* genus, including species found in the Mediterranean (*A. arborescens*, *A. caerulescens* subsp., and *A. annua*), are rich in the majority of these components. Other regions that produce EOs

include North America, Turkey, India, and France [26, 27]. Also, EOAA and the chemicals found in essential oils of the Moroccan-endemic *A. mesatlantica* are somewhat similar. It follows that EOAA's high concentration of artemisia ketone may account for its antioxidant power. According to the literature and the DPPH test, artemisia ketone is more active than camphor and 1,8-cineole. Research has shown that the active ingredients in essential oils are often responsible for their antioxidant properties [28-30]. The antioxidant powder of essential oils is determined by the synergistic action of their minor components. When comparing OEs with and without hydrocarbon terpenes, those rich in oxygenated monoterpenes tend to have stronger antiradical effects. The antioxidant results align with earlier research that indicated *Artemisia* to have promising antioxidant activity across all bioassays, including DPPH,  $\beta$ -carotene bleaching, and total antioxidant capacity. This is an important finding. At the moment, scientists are trying to figure out how artemisinin, the active ingredient in *A. annua* L., works to prevent malaria. Additionally, *A. annua* L. has been the subject of research into its possible medicinal benefits for a wide range of illnesses in the last several decades. These include inflammatory and malignant disorders [31], infections caused by bacteria, viruses, and parasites, and more. According to our research, the antibacterial activity of the ethanolic *A. annua* extract was greater.

## CONCLUSION:

Antibacterial activity against *Staphylococcus aureus*, *Escherichia coli*, *Streptococcus pyogenes*, and *Staphylococcus epidermidis* was demonstrated by means of the polyphenolic compounds that are prevalent in the leaves of the *Artemisia annua* L. species. The level of efficiency against *Staphylococcus aureus* ( $28.00 \pm 0.45$ ) was much higher when *Artemisia annua* was used. Based on our findings, it is worth further investigation to explore the possibilities of using *Artemisia* extracts in new therapy regimens for infectious disorders that are resistant to current treatments. *Artemisia annua* extracts showed significant antibacterial action against multidrug-resistant *Escherichia coli*, according to the study, which provides hope for a natural substitute for traditional antibiotics and food preservatives. Natural extracts from *Artemisia* may play a pivotal role in creating new treatments and preventative measures against multidrug-resistant bacteria, providing a sustainable and environmentally friendly answer to the problem of antibiotic resistance. These results provide strong evidence in favor of additional research into the therapeutic use of *Artemisia* extracts against resistant infections.

## REFERENCES

1. Batiha, G.E.; Olatunde, A.; El-Mleeh, A.; Hetta, H.F.; Al-Rejaie, S.; Alghamdi, S.; Zahoor, M.; Magdy Beshbishy, A.; Murata, T.; Zaragoza-Bastida, A.; et al. Bioactive Compounds, Pharmacological Actions, and Pharmacokinetics of Wormwood (*Artemisia absinthium*). *Antibiotics* 2020, 9, 353.
2. Mamatova, A, Korona-Glowniak, I. Phytochemical composition of wormwood (*Artemisia gmelinii*) extracts in respect of their antimicrobial activity. *BMC Complement. Altern. Med.* 2019, 19, 288.
3. AlSalhi, M.S.; Elumalai, K.; Devanesan, S.; Govindarajan, M.; Krishnappa, K.; Maggi, F. The aromatic ginger *Kaempferia galanga* L.(Zingiberaceae) essential oil and its main compounds are effective larvicidal agents against *Aedes vittatus* and *Anopheles maculatus* without toxicity on the non-target aquatic fauna. *Ind. Crops Prod.* 2020, 158, 113012.
4. El Moussaoui, A.; Bourhia, M.; Jawhari, F.Z.; Salamatullah, A.M.; Ullah, R.; Bari, A.; Majid Mahmood, H.; Sohaib, M.; Serhii, B.; Rozhenko, A.; et al. Chemical Profiling, Antioxidant, and Antimicrobial Activity against Drug-Resistant Microbes of Essential Oil from *Withania frutescens* L. *Appl. Sci.* 2021, 11, 5168.
5. Chebbac, K.; Moussaoui, A.E.; Bourhia, M.; Salamatullah, A.M.; Alzahrani, A.; Guemmouh, R. Chemical analysis and antioxidant and antimicrobial activity of essential oils from *Artemisia negrei* L. Against drug-resistant microbes. *Evid. -Based Complement. Altern. Med.* 2021, 2021, 5902851.
6. Chaachouay, N.; Douira, A.; Hassikou, R.; Brhadda, N.; Dahmani, J.; Belahbib, N.; Ziri, R.; Zidane, L. Mr Chaachouay Nouredine Sous le thème" Etude floristique et ethnométriciale des plantes aromatiques et médicinales dans le Rif (Nord du Maroc). Ph.D. Thesis, Département de Biologie-Université Ibn Tofail-Kénitra, Kenitra, Morocco, 2020.

7. Tahri, N.; El Basti, A.; Zidane, L.; Rochdi, A.; Douira, A. Etude ethnobotanique des plantes medicinales dans la province de Settat (Maroc). *Kast. Univ. J. For. Fac.* 2012, 12, 192–208.
8. Yuan, H.; Ma, Q.; Ye, L.; Piao, G. The traditional medicine and modern medicine from natural products. *Molecules* 2016, 21, 559.
9. Gruessner, B.M.; Cornet-Vernet, L.; Desrosiers, M.R.; Lutgen, P.; Towler, M.J.; Weathers, P.J. It is not just artemisinin: *Artemisia* sp. for treating diseases including malaria and schistosomiasis. *Phytochem. Rev.* 2019, 18, 1509–1527.
10. Nigam, M.; Atanassova, M.; Mishra, A.P.; Pezzani, R.; Devkota, H.P.; Plygun, S.; Salehi, B.; Setzer, W.N.; Sharifi-Rad, J. Bioactive compounds and health benefits of *Artemisia* species. *Nat. Prod. Commun.* 2019, 14, 1934578X19850354.
11. Abad, M.J.; Bedoya, L.M.; Apaza, L.; Bermejo, P. The *Artemisia* L. genus: A review of bioactive essential oils. *Molecules* 2012, 17, 2542–2566.
12. Jaradat, N.; Qneibi, M.; Hawash, M.; Al-Maharik, N.; Qadi, M.; Abualhasan, M.N.; Ayeshe, O.; Bsharat, J.; Khadir, M.; Morshed, R.; et al. Assessing *Artemisia arborescens* essential oil compositions, antimicrobial, cytotoxic, anti-inflammatory, and neuroprotective effects gathered from two geographic locations in Palestine. *Ind. Crops Prod.* 2022, 176, 114360.
13. Hinane, D.; Oubaha, S.; Satrani, B.; Ghanmi, M.; Bourkhiss, B. Domestication of the endemic plant of Morocco and high added value: *Artemisia herba alba*. *J. Mater. Environ. Sci.* 2020, 11, 283–292.
14. Farzaneh, F.; Ebrahim, H.S.; Akbar, V. Investigating on Effect of Wormwood Extract on Reduction of Renal Toxicity in Treated Rats by Azathioprine. *Biomed. Pharmacol. J.* 2015, 8, 291–299.
15. Kim, M.H.; Seo, J.Y.; Liu, K.H.; Kim, J.S. Protective effect of *Artemisia annua* L. extract against galactose-induced oxidative stress in mice. *PLoS ONE* 2014, 9, e101486.
16. Fiamegos, Y.; Kastritis, P.; Exarchou, V. Antimicrobial and efflux pump inhibitory activity of caffeoylquinic acids from *Artemisia absinthium* against gram-positive pathogenic bacteria. *PLoS ONE* 2011, 6, e18127.
17. Hori R, Sugiyama J. A combined FTIR microscopy and principal component analysis on softwood cell walls. *Carbohydr Polym.* 2003;52:449–453.
18. Semeniuc, C, Rotar, A. Antibacterial activity and interactions of plant essential oil combinations against Grampositive and Gram-negative bacteria. *J. Food Drug Anal.* 2017, 25, 403–408.
19. Qian, W.; Yang, M.; Wang, T.; Sun, Z.; Liu, M.; Zhang, J.; Zeng, Q.; Cai, C.; Li, Y. Antibacterial Mechanism of Vanillic Acid on Physiological, Morphological, and Biofilm Properties of Carbapenem-Resistant *Enterobacter hormaechei*. *J. Food Prot.* 2020, 83, 576–583.
20. Sethupathy, S.; Ananthi, S.; Selvaraj, A.; Shanmuganathan, B.; Vigneshwari, L.; Balamurugan, K.; Mahalingam, S.; Pandian, S.K. Vanillic acid from *Actinidia deliciosa* impedes virulence in *Serratia marcescens* by affecting S-layer, flagellin and fatty acid biosynthesis proteins. *Sci. Rep.* 2017, 7, 16328.
21. Yemis, G.P.; Pagotto, F.; Bach, S.; Delaquis, P. Effect of vanillin, ethyl vanillin, and vanillic acid on the growth and heat resistance of *Cronobacter* species. *J. Food Prot.* 2011, 74, 2062–2069.
22. Mamatova, A.S.; Korona-Głowniak, I.; Skalicka-Wozniak, K.; Jozefczyk, A.; Wojtanowski, K.K.; Baj, T.; Sakipova, Z.B.; Malm, A. Phytochemical composition of wormwood (*Artemisia gmelinii*) extracts in respect of their antimicrobial activity. *BMC Complement. Altern. Med.* 2019, 19, 288.
23. Masoud, S. M., Abd El-Baky, R. M., Aly, S. A., & Ibrahem, R. A. (2021). Co-existence of certain ESBLs, MBLs and plasmid mediated quinolone resistance genes among MDR *E. coli* isolated from different clinical specimens in Egypt. *Antibiotics*, 10(7), 835.
24. Mathlouthi, A., Saadaoui, N., & Ben-Attia, M. (2021). Essential oils from *Artemisia* species inhibit biofilm formation and the virulence of *Escherichia coli* EPEC 2348/69. *Biofouling*, 37(2), 174-183.
25. Navarro-Pérez, M. L., Vadillo-Rodríguez, V., Fernández-Babiano, I., Pérez-Giraldo, C., & Fernández-Calderón, M. C. (2021). Antimicrobial activity of a novel Spanish propolis against planktonic and sessile oral *Streptococcus* spp. *Scientific Reports*, 11(1), 23860.
26. Poiata, A., Tuchilus, C., Ivanescu, B., Ionescu, A., & Lazar, M. I. (2009). Antibacterial activity of some *Artemisia* species extracts. *Revista MedicoChirurgicală a Societății de Medici și Naturaliști din Iași*, 113, 911-914.

27. Salmerón-Manzano, E., Garrido-Cardenas, J. A., & Manzano-Agugliaro, F. (2020). Worldwide research trends on medicinal plants. *International Journal of Environmental Research and Public Health*, 17(10), 3376.
28. Sengul, M., Ercisli, S., Erzurum, T., Yildiz, H., Gungor, N., Kavaz, A., & Çetin, B. (2011). Antioxidant, antimicrobial activity, and total phenolic content within the aerial parts of *Artemisia absinthum*, *Artemisia santonicum*, and *Saponaria officinalis*. *Iranian Journal of Pharmaceutical Research*, 10, 49-56.
29. Shaaban, M. T., Abdel-Hamid, M. S., Orabi, S. H., Korany, R. M., & Elbawab, R. H. (2024). Assessment of the antibacterial efficacy of silver nanoparticles-based *Artemisia annua* against methicillin-resistant *Staphylococcus aureus* infected lung tissues in albino rats. *Journal of Analytical Science and Technology*, 15(1), 25.
30. Shaaban, M. T., El Silk, S. E., & Tayel, M. A. (2011). Efficiency of some plant extracts, carbohydrates and inorganic salts as anti-adhesion agents against the adhesion of *Staphylococcus* strains to HEp-2 cells. *Life Sci J*, 8, 1172-1182.
31. Shaaban, M. T., Ibrahim, H. A. H., & Hanafi, A. A. M. (2020). Antibiotic-resistant bacteria isolated from selected urine and stool human specimens. *Biosci Res*, 17(1), 351-365.